

MINOR AND TRACE STEROLS IN MARINE INVERTEBRATES 30.<sup>1</sup> ISOLATION, STRUCTURE  
ELUCIDATION AND PARTIAL SYNTHESIS OF 26-METHYLSTRONGYLOSTEROL AND 28-METHYLXESTOSTEROL  
- TWO MARINE STEROLS ARISING BY A NOVEL QUADRUPLE BIOMETHYLATION SEQUENCE

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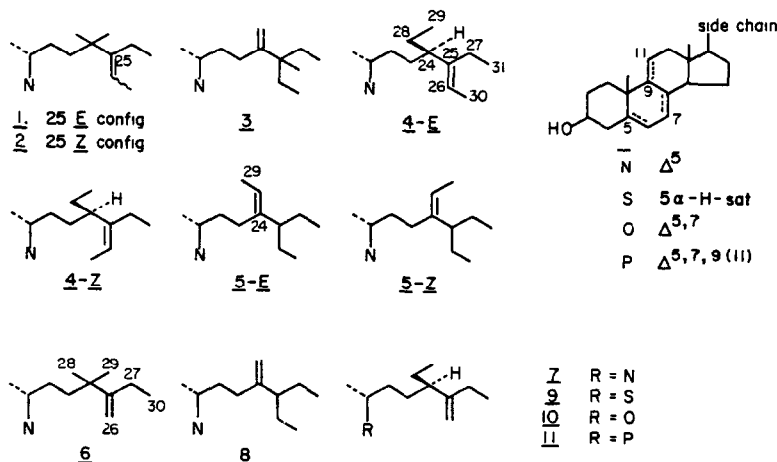
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Abstract: Two new sterols, 26-methylstrongylosterol (4) and 28-methylxestosterol (5), arising from a hitherto unknown quadruple biomethylation sequence, together with the biogenetically important "missing link", durissimasterol (6), were isolated from the Indo-Pacific sponge *Strongylophora durissima*.

Recently we documented the first examples of quadruple biomethylation in the sterol side chain - xestospongesterol (1),<sup>3</sup> isoxestospongesterol (2)<sup>3</sup> and 25-methylxestosterol (3).<sup>4</sup> The eventual isolation of a sterol resulting from an alternative quadruple biomethylation sequence at positions 24, 26, 27 and 28 has been predicted earlier<sup>5</sup> and one such candidate - 24R-ethyl-26, 27-dimethylcholesta-5,25(26)dien-3 $\beta$ -ol (26-methylstrongylosterol) (4-Z) - was synthesized.<sup>5</sup> We now wish to report the first occurrence of two sterols, 26-methylstrongylosterol (4) and 28-methylxestosterol (5), arising from such a quadruple biomethylation pattern together with a biogenetically important "missing link",<sup>6</sup> now named durissimasterol (6), in the Indo-Pacific sponge *Strongylophora durissima*, in which the main sterol is strongylosterol (7).<sup>7</sup>

The mass spectrum of 26-methylstrongylosterol (4) ( $M^+$  440) was identical with that of the synthetic<sup>5</sup> Z isomer (4-Z), but the doubling of the relevant 360 MHz proton NMR peaks (Table 1) indicated that the natural sterol was a mixture of E and Z isomers of 4. Verification was provided by synthesis via a known procedure,<sup>5</sup> using benzene instead of THF<sup>5</sup> in the Wittig condensation. Reverse-phase HPLC yielded 4-E [mp 90-91° C,  $[\alpha]_D^{20}$  -21° (c, 0.002, CHCl<sub>3</sub>)] and 4-Z [mp 107-108° C,  $[\alpha]_D^{20}$  -15° (c, 0.017, CHCl<sub>3</sub>)], both of which had the same GC mobility (rrt 2.26; cholesterol=1.00) as the natural compound. The C-26 and C-31 proton signals were more shielded in the E isomer,<sup>8</sup> (Table 1) while a septet at 2.475 ppm (C-24 allylic proton) was observed in the Z isomer. Final proof of the stereochemical assignment was provided by the <sup>13</sup>C NMR spectra, in which an upfield shift of 7.8 ppm and a downfield shift of 5.2 ppm respectively were observed for C-24 and C-26 in the Z isomer.<sup>9</sup>

The mass spectra of the second new sterol, 28-methylxestosterol (5) ( $M^+$  440), and of xestosterol (8) ( $M^+$  426) were practically identical below their  $m/z$  314 base peak and showed virtually no peaks in the region  $M^+ \rightarrow m/z$  314, thus pointing to the presence of a  $\Delta^{24(28)}$ -double bond.<sup>11</sup> Attachment of a methyl group to the  $\Delta^{24(28)}$ -double bond of 5 was deduced from the NMR (Table 1) doublet at 1.679 ppm, which was coupled to the olefinic proton at 5.25 ppm, while the six-proton triplet at 0.930 ppm indicated the presence of a 30,31-dimethyl moiety. The stereo-



chemistry of the  $\Delta^{24(28)}$ -double bond was proven by comparison with the synthetic isomers  $5-E$  and  $5-Z$ , obtained by Wittig condensation of 26,27-dimethyl-6 $\beta$ -methoxy-3 $\alpha$ ,5-cyclocholestan-24-one<sup>10</sup> with triphenylethylphosphonium bromide, followed by removal of the protecting group [ $5-E$ : mp 116-117°C,  $[\alpha]_D^{20}$  -37° (c, 0.009, CHCl<sub>3</sub>);  $5-Z$ : mp 130-131°C,  $[\alpha]_D^{20}$  -15° (c, 0.02, CHCl<sub>3</sub>)] and comparison of their NMR spectra (notably a pentet at 2.40 ppm (C-25H) in  $5-Z$  vs. 1.8 ppm in  $5-E$ ) with those<sup>12</sup> of isofucosterol ( $Z$  isomer, C-25 H at 2.8 ppm) and fucosterol ( $E$  isomer, C-25 H at 2.2 ppm). This stereochemical assignment was confirmed by the <sup>13</sup>C NMR spectra, in which an upfield shift of 7.76 ppm for C-25 was observed in the  $Z$  isomer. The NMR spectra (Table 1) of synthetic and natural  $5-E$  were identical as were their mass spectra and GC rrt (2.26). While  $4$  and  $5$  possess the same GC rrt, they can be distinguished by reverse-phase HPLC (Whatman Partisil M9 10/50 ODS-2 column) rrt:1.20 ( $4$ ) vs. 1.12 ( $5$ ).

The third new sterol, named durissimasterol ( $6$ ) ( $M^+$  426), showed typical peaks ( $m/z$  213, 231, 253, 271) of a  $\Delta^5$ -3 $\beta$ -hydroxy sterol,<sup>13</sup> an intense peak at  $m/z$  328 (typical<sup>11,14</sup> of a McLafferty rearrangement in a  $\Delta^{25}$ -unsaturated side chain), and a base peak at  $m/z$  98 (C<sub>7</sub>H<sub>14</sub> due to "reverse" McLafferty rearrangement); the absence of a  $m/z$  314 peak suggested the presence of a C-24 quaternary center.<sup>3</sup> The NMR spectrum (Table 1) pointed directly to structure  $6$ , which is an important "missing link" in the proposed<sup>3,6</sup> biosynthesis of (iso)xestospongesterol ( $1,2$ ). For confirmation,  $6$  [mp 142-143°C,  $[\alpha]_D^{20}$  -36° (c, 0.006, CHCl<sub>3</sub>)] was synthesized by Wittig condensation of the appropriate ketone (obtained as a by-product in an earlier<sup>3</sup> synthesis) with triphenylmethylphosphonium bromide (*n*-BuLi, THF, 4 hr, reflux), followed by deprotection of the *i*-methyl ether. The NMR (Table 1) and mass spectra, as well as GC rrt (1.91) of the natural and synthetic sterols, were identical.

Three new minor sterols with  $M^+$  428, 424 and 422 were identified as strongylostanol ( $9$ ), 7-dehydrostrongylosterol ( $10$ ) and 7,9(11)-didehydrostrongylosterol ( $11$ ) by comparison of their 360 MHz proton NMR (identical side chain proton signals) and mass spectra with those of strongylosterol ( $7$ ). Thus the high resolution mass spectrum of strongylostanol ( $9$ ) ( $M^+$  428.4037) dis-

played the diagnostic peaks ( $m/z$  215,233,255 and 273) for the nucleus of a saturated stanol. The absence of a C-6 vinylic proton in its NMR spectrum and the expected<sup>15</sup> chemical shifts for the C-18 and C-19 methyl groups (0.631 and 0.797 ppm) were only consistent with a 3 $\beta$ -hydroxy-5 $\alpha$ -saturated nucleus. The nature of the extra unsaturation in the nuclei of 10 and 11 was demonstrated easily by the characteristic mass and NMR spectra of such  $\Delta^{5,7}$  and  $\Delta^{5,7,9(11)}$ -unsaturated 3 $\beta$ -hydroxy sterols.<sup>16,17</sup>

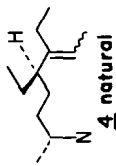
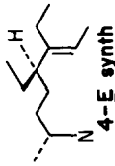
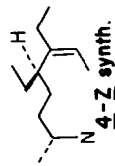
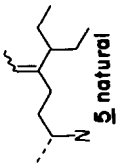
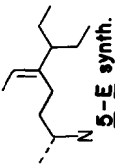
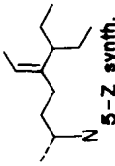
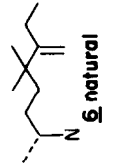
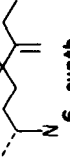
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Table 1. Selected  $^1\text{H}$  360 MHz NMR Chemical Shifts ( $C_6D_6$ ) of 26-Methylstrongylosteron (4), 28-Methylxestosteron (5) and Durissimasteron (6).

	C-18 Me	C-19 Me	C-21 Me	C-29 Me	C-30 Me	C-31 Me	C-26 H	C-28 H
 4-natural	0.659 (48%) 0.666 (52%)	0.941	1.027 (d, J=6.4) 1.034 (d, J=6.2)	0.885 (t, J=7.4) 0.920 (t, J=7.4)	1.665 (d, J=6.7) 1.625 (d, J=6.7)	1.074 (t, J=7.4) 1.025 (t, J=7.4)	5.44 5.24	
 4-E synth	0.666	0.940	1.034 (d, J=6.2)	0.919 (t, J=7.5)	1.626 (d, J=6.7)	1.025 (t, J=7.5)	5.254 (q, J=6.7)	
 4-Z synth.	0.658	0.941	1.026 (d, J=6.5)	0.884 (t, J=7.4)	1.663 (d, J=6.7)	1.074 (t, J=7.4)	5.444 (q, J=6.7)	
 5-natural	0.664	0.949	1.060 (d, J=6.7)	1.679 (d, J=6.6)	0.930 (t, J=7.5)	0.930 (t, J=7.5)	5.250 (q, J=6.3)	
 5-E synth.	0.664	0.949	1.060 (d, J=6.5)	1.678 (d, J=6.7)	0.929 (t, J=7.3)	0.929 (t, J=7.3)	5.250 (q, J=6.8)	
 5-Z synth.	0.660	0.942	1.036 (d, J=6.5)	1.645 (d, J=6.8)	0.905 (t, J=7.4)	0.905 (t, J=7.4)	5.530 (q, J=6.7)	
 6-natural	0.657	0.944	0.990 (d, J=6.5)	1.088 <sup>a</sup>		1.077 (t, J=7.4)	4.98 4.94	
 6 synth.	0.658	0.944	0.990 (d, J=6.5)	1.088 <sup>a</sup>		1.077 (t, J=7.4)	4.978 4.94	

<sup>a</sup> Six-proton signal due to C-28 and C-29 methyl groups.